Nordic immunohistochemical Quality Control

Institute of Pathology, Aalborg University Hospital, Ladegaardsgade 3, 9000 Aalborg, Denmark

Scheme 2024, Assessment Run H26

Aalborg, 2nd September 2024

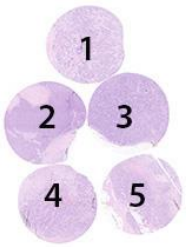
Dear participant

Pairs of unstained slides of NordiQC multi-tissue samples have been sent according to your submitted protocols for this run.
If you are in any doubt, please read the details on the website: www.nordiqc.org, or send us an e-mail: nordiqc@rn.dk. Also inform us if slides are missing.

Please note your participant number (NQC…) shown on the address label. This number **must** be indicated on all slides and used for any communication with NordiQC.

Contents of the NordiQC multi-tissue samples

Top row closest to the glass label.



HER-2 ISH:
1-5: Breast carcinoma

For each marker stain the sections according to the protocols you have uploaded on our website. **If any changes are made in the protocols, upload the used protocols as these are used for data analysis and individual feed-back in case needed.**

In-house controls shall, if possible, be placed on the NordiQC slide (see next page).

Score the slide using the electronic scoring sheet on the web site (login and choose “scoring sheet” in the “options”-menu on the right).

HER-2 FISH: Do not return the FISH-stained slide to NordiQC.

HER-2 BRISH: Mark the slides with your NordiQC participant number and return one stained slide to NordiQC. Keep the remaining NordiQC slide to serve as a control set.

Do not return separate in-house controls.

Deadline for slide return: 10th October.

Slides return address:
NordiQC
Institute of Pathology
Aalborg University Hospital
Ladegaardsgade 3
9000 Aalborg
Denmark

Overall and individual results are available on the website on 10th December.

Thank you for your assistance.

Best regards
NordiQC

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General information;

NordiQC tissue material and sections:
All included tissues in the NordiQC multi-tissue samples have been fixed in 10% neutral buffered formalin and embedded in paraffin.

The sections of the multi-tissue blocks are sectioned at 3-4 µm.

After sectioning, the slides are air dried at room temp. for 48 hours, after which they are stored at -80°C until distributed.

After receipt of slides, store them at -20°C until staining.

The slides are unbaked.

Before staining in-house control material should be mounted on the NordiQC slide and dried at e.g. 37°C overnight or 55-60°C for one hour to secure section adhesion.

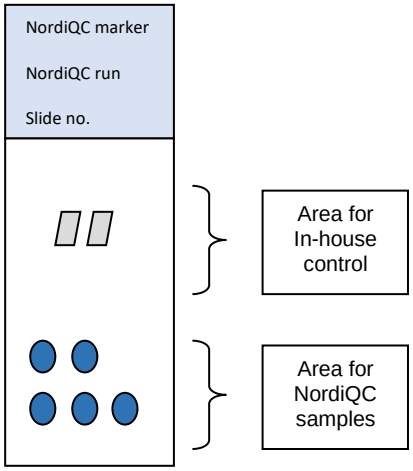

Fig. 1 Example of NordiQC slide

BRISH staining
The NordiQC slides/samples should be stained in the same manner as the routine samples for the specific markers and should not be handled differently.

Slide return
Return slides to NordiQC after BRISH staining has been completed and in due time for deadlines indicated in this accompany letter and at www.nordiqc.org/modules.
The slides must be returned in the same or a similar box in which it was received.

You can see if NordiQC has received your slides. Please log in at our website and choose “Protocol submission”. When the slides are received at NordiQC and registered, this will be marked with the date received in the column “Slide received by NordiQC”.

Submitted sections
Submitted sections are stored by NordiQC for future documentation and will not be returned to the laboratories. However, laboratories can request their stained sections for review e.g. in case their own control set is lost.

In-house on-slide controls
In-house on-slide controls shall serve as participants internal IHC system control to verify successful test. In-house on-slide controls will not be evaluated by NordiQC as expected patterns and content of tissue are not known and validated by NordiQC. The results in the controls will not affect the final individual assessment mark.

Results
The overall assessment results for all markers are published in reports available at www.nordiqc.org/epitope at the time indicated at www.nordiqc.org/modules.

The assessment reports will provide an overview of assessment criteria, protocols, reagents and instruments used by the participants and results obtained. The report will provide guidelines for recommendable control materials and descriptions / illustrations of the expected staining patterns in these.

In case of an insufficient result assessed as borderline or poor, the participant will receive individual suggestions for protocol improvement.